

## Genotyping Protocol: **MMRRC 442**

**Assay Type:** PCR (can distinguish heterozygous animals from homozygous animals) detects presence of mutations in both the *Chrna5* ( $\alpha 5$ ) and *Chrb4* ( $\beta 4$ ) genes.

**DNA Extraction:** DNA from tail snips was extracted using Sigma's Extract-N-Amp Tissue PCR Kit (Cat#XNAT2R). Kit directions for fresh or frozen tails were performed with a few minor modifications as follows: use 50  $\mu$ l of Extraction Solution and 12.5  $\mu$ l of Tissue Preparation Solution and 50  $\mu$ l of Neutralization Solution B.

**Strain Information:** A neo cassette replaces most of Exon 5 in the *Chrna5* gene. Most of Exon 5 is deleted in the *Chrb4* gene. Details can be found in Kedmi et al (2004) *Physiol. Genomics* 17:221-229.

### **A5 Primer Information:**

- |                  |   |
|------------------|---|
| 1) Name: A5 Wt f | Sequence: 5'- GTG AAA GAG AAC GAC GTC CGC -3'     |
| 2) Name: A5 Wt R | Sequence: 5'- GCC TCA GCC CCT GAA TGG TAG -3'     |
| 3) Name: Neo-1   | Sequence: 5'- CTT TTT GTC AAG ACC GAC CTG TCC -3' |
| 4) Name: Neo-2   | Sequence: 5'- CTC GAT GCG ATG TTT CGC TTG GTG -3' |

### **B4 Primer Information:**

- |                     |  |
|---------------------|--|
| 1) Name: B4 Rev.wt  | Sequence: 5'-AGTGAATGACAGATGGACCTC -3' |
| 2) Name: B4 Rev.mut | Sequence: 5'- AGTCAAAACCTTGCACCAG -3'  |
| 3) Name: B4 Forward | Sequence: 5'- AGTCAAAACCTTGCACCAG -3'  |

**Assay names: A5 PCR, B4 PCR**

### **PCR Master Mix Components:**

#### **Master Mix for A5 assay:**

Component	Manufacturer	Concentration	$\mu$ l/rxn
Extract-N-Amp Reaction Mix	Sigma (Cat#XNAT2R)	2X	10
A5 Wt F	Sigma-Genosys	25 $\mu$ l	0.3
A5 Wt R	Sigma-Genosys	25 $\mu$ l	0.3
Neo-1	Sigma-Genosys	25 $\mu$ l	0.3
Neo-2	Sigma-Genosys	25 $\mu$ l	0.3
water			4.8

#### **Master Mix for B4 assay:**

Component	Manufacturer	Concentration	$\mu$ l/rxn
Extract-N-Amp Reaction mix	Sigma	2X	10
B4 Rev.wt	Sigma-Genosys	25 $\mu$ l	0.3
B4 Rev.mut	Sigma-Genosys	25 $\mu$ l	0.3
B4 Forward	Sigma-Genosys	25 $\mu$ l	0.3
water			5.1

### **PCR Setup:**

Both assays final reaction: 16 $\mu$ l master mix & 4 $\mu$ l DNA template (10-20 ng/ $\mu$ l DNA)  
All reactions were performed in 200 $\mu$ l thin walled PCR tubes and were run in an Applied Biosystems 2700 thermocycler.

### **Cycle Parameters (Both Assays):**

- |    |      |           |
|----|------|-----------|
| 1) | 94°C | 3 minutes |
| 2) | 94°C | 1 minute  |
| 3) | 58°C | 1 minute  |

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08.01.16 MLS

- 4) 72°C 1 minute
- 5) Repeat steps 2-4 34 times for a total of 35 cycles
- 6) 72°C 10 minutes
- 7) 4°C hold until refrigerate product

**Product Analysis:**

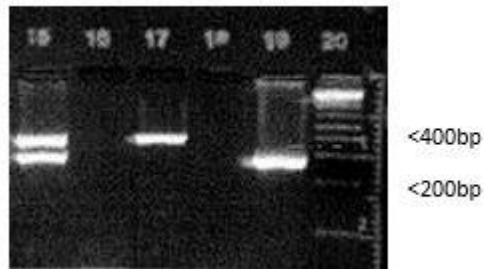
All products were analyzed on a 3% agarose gel with ethidium bromide staining

**Expected products:**

- A5 assay Hom mutant = 290 bp  
WT = 380 bp  
Het = 290 and 380 bp
  
- B4 assay Hom mutant = 150 bp  
WT = 300 bp  
Het = 150 and 300 bp

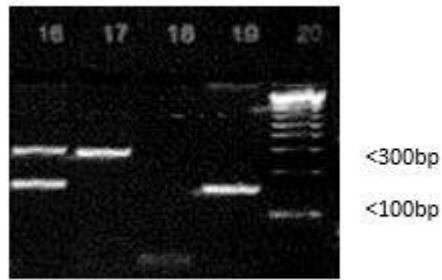
**Example gels:**

**A5 Assay:**



Lane 15 displays a heterozygous sample (290bp and 380bp products)  
Lane 17 displays a WT sample (380bp product)  
Lane 19 displays a homozygous mutant sample (290bp product)  
Lanes 16 and 18 are PCR blanks  
Lane 20 is 1kb+ Ladder (Invitrogen Cat#10787-018)

**B4 Assay:**



Lane 16 displays a heterozygous sample (150bp and 300bp products)  
Lane 17 displays a WT sample (300bp product)  
Lane 19 displays a homozygous mutant sample (150bp product)  
Lane 18 is a PCR blank  
Lane 20 is 1kb+ Ladder (Invitrogen Cat#10787-018)